

Toward Improved Influenza Vaccines

Stephen J. Kent

Department of Microbiology and Immunology, University of Melbourne, Australia

(See the major article by Carroll et al 24–33.)

Keywords. Influenza; vaccine; adjuvant; immunity; antibodies.

Influenza takes a large toll on human health, particularly in the elderly population. The current unadjuvanted trivalent inactivated influenza vaccines (TIVs) are notorious for modest-to-dismal efficacy among elderly individuals. Protection by TIV is limited largely to the 3 or 4 strains chosen for each influenza season. The drifts in circulating strains mean that annual receipt of TIV is recommended, the only licensed vaccine with such frequent administration. Furthermore, this current strategy provides no cross-protection against highly novel pandemic influenza viruses. There is a delay between identifying a pandemic strain to use in a vaccine and generating a matching vaccine, leaving the population highly vulnerable for some months. A considerable worldwide effort is underway to generate much improved influenza vaccines, particularly for the most vulnerable populations.

The study by Carroll et al in this issue of the *Journal* is timely research in this regard [1]. The group studied a cohort of elderly macaques that were aged >18 years. Similar to elderly humans, these elderly macaques had reduced levels of

total T cells, especially naive CD4⁺ T cells. The addition of a novel adjuvant to 2 doses of a standard TIV dramatically improved the immunogenicity and protective efficacy of the TIV. A liposome/DNA adjuvant, cationic lipid/DNA complex (CLDC), appeared safe and induced higher and more durable levels of protective antibodies, compared with the unadjuvanted TIV. Antibody levels in the elderly macaques that received TIV with the adjuvant approached those seen previously in juvenile macaques given unadjuvanted TIV [2]. There was clear evidence of much better clearance of a challenge influenza virus in the elderly macaques that received the novel adjuvanted TIV, compared with those that received the standard TIV.

Why did the adjuvant work? The levels of influenza virus–neutralizing (hemagglutination-inhibiting) antibodies induced were robust, and the challenge used a strain that closely matched that in the vaccines, as would occur if the seasonal TIV strains matched the circulating infecting strains. However, the adjuvant also induced some CD4⁺ and CD8⁺ T-cell immunity that may have assisted the protective efficacy [3]. T-cell immunity is often directed to conserved internal antigens that can provide cross-protection against divergent strains. It is also possible that the CLDC-adjuvanted TIV generated functional nonneutralizing antibodies, such as those involved in antibody-dependent cellular cytotoxicity, that may also have assisted in the protective

immunity observed [4]. Such antibodies, like T cells, are often cross-reactive to divergent strains and may provide some cross-protection [5,6]. These issues could be teased out with passive transfer or heterologous challenge studies in the future.

The translation of these promising macaque results to humans remains to be shown. A key issue in clinical trials will be the reactogenicity of the adjuvant. Although no marked effects were observed in this macaque study, macaques tend not to complain much, and both the general population and regulatory bodies are woefully intolerant of even modest reactogenicity. This is one reason our current TIV remains unadjuvanted. A surprising proportion of humans even fervently believe the current TIV can cause influenza.

A second key issue for clinical trials will be the humoral immunogenicity of CLDC-adjuvanted influenza vaccines. There is reason to be hopeful that results of human studies will be positive. The elderly captive macaques studied (age, 18–22 years) will have had much less prior exposure to influenza viruses than elderly humans (ie, those aged >65 years). It may well be even easier for such vaccines boost low-level preexisting anti-influenza virus immune responses induced by prior infection in humans than it was to generate new responses in macaques. Indeed, recent evidence suggests that even current unadjuvanted TIVs primarily boost low-level preexisting influenza virus antibody responses rather than generate new responses [7]. Interestingly, the

Received 25 July 2013; accepted 25 July 2013; electronically published 17 October 2013.

Correspondence: Stephen J. Kent, MD, Department of Microbiology and Immunology, University of Melbourne 3010 Australia (skent@unimelb.edu.au).

The Journal of Infectious Diseases 2014;209:4–5

© The Author 2013. Published by Oxford University Press on behalf of the Infectious Diseases Society of America. All rights reserved. For Permissions, please e-mail: journals.permissions@oup.com.

DOI: 10.1093/infdis/jit542

clear superiority of the CLDC-adjuvanted vaccine was not demonstrated until following the second dose, which may be necessary in future clinical studies.

Where does this CLDC adjuvant approach fit with other efforts at developing better influenza vaccines? Adjuvanted influenza vaccines have now reached the market, particularly the squalene-based oil-in-water adjuvants from GlaxoSmith-Kline (AS03), Sanofi Pasteur (AF03), and Novartis (MF59). These vaccines generally show important incremental improvements in immunogenicity with modest increases in reactogenicity, compared with unadjuvanted TIVs [8].

There is substantial current research investment in developing influenza vaccines with quantum leaps in efficacy, durability, and cross-protection capacity over the current vaccines. Novel live-attenuated virus approaches look promising in animal models [9]. Lipid-based virus-like particles (“virosomes”) [10] and nanoparticle-based approaches show considerable efficacy in early studies [11]. Recent research shows the promise of using vectors to deliver the rare neutralizing antibodies that cross-recognize divergent influenza virus strains [12]. Various viral vector vaccines, designed, in part, to induce T-cell immunity, also show promise [13]. The role of T-cell immunity in controlling immunity will be interesting—it could be argued that T-cell responses may expand too late to be of use in early control of acute influenza virus replication. However, there is solid

evidence that T-cell immunity plays a role in primate models [14], and in humans T-cell immunity could play an important role in controlling severe protracted influenza virus infections, which are associated with much of the influenza-associated morbidity and mortality.

Overall, there is a great need to develop better influenza vaccines for the elderly population and other vulnerable groups. The work described by Carroll et al describes another promising approach moving toward human efficacy trials.

Notes

Financial support. This work was supported by the Australian National Health and Medical Research Council (awards 510448, 1042634, and 1041832).

Potential conflicts of interest. Author certifies no potential conflicts of interest.

The author has submitted the ICMJE Form for Disclosure of Potential Conflicts of Interest. Conflicts that the editors consider relevant to the content of the manuscript have been disclosed.

References

1. Carroll TD, Matzinger SR, Barry PA, McChesney MB, Fairman J, Miller CJ. Effectiveness of influenza vaccination of elderly rhesus macaques is dramatically improved by addition of a cationic lipid/DNA adjuvant. *J Infect Dis* **2014**; 209:24–33.
2. Carroll TD, Matzinger SR, Barro M, et al. Alphavirus replicon-based adjuvants enhance the immunogenicity and effectiveness of Fluzone (R) in rhesus macaques. *Vaccine* **2011**; 29:931–40.
3. Sun J, Braciale TJ. Role of T cell immunity in recovery from influenza virus infection. *Curr Opin Virol* **2013**; 3:425–9.
4. Jegaskanda S, Weinfurter JT, Friedrich TC, Kent SJ. Antibody-dependent cellular cytotoxicity is associated with control of

5. Jegaskanda S, Job ER, Kramski M, et al. Cross-reactive influenza-specific antibody-dependent cellular cytotoxicity antibodies in the absence of neutralizing antibodies. *J Immunol* **2013**; 190:1837–48.
6. Jegaskanda S, Laurie K, Amarasena T, et al. Age-associated cross-reactive ADCC toward 2009-pandemic influenza. *J Infect Dis* **2013**; 208:1051–61.
7. Li GM, Chiu C, Wrammert J, et al. Pandemic H1N1 influenza vaccine induces a recall response in humans that favors broadly cross-reactive memory B cells. *Proc Natl Acad Sci U S A* **2012**; 109:9047–52.
8. Nicholson KG, Abrams KR, Batham S, et al. Immunogenicity and safety of a two-dose schedule of whole-virion and AS03A-adjuvanted 2009 influenza A (H1N1) vaccines: a randomised, multicentre, age-stratified, head-to-head trial. *Lancet Infect Dis* **2011**; 11:91–101.
9. Hatta Y, Hatta M, Bilsel P, Neumann G, Kawaoka Y. An M2 cytoplasmic tail mutant as a live attenuated influenza vaccine against pandemic (H1N1) 2009 influenza virus. *Vaccine* **2011**; 29:2308–12.
10. Radosevic K, Rodriguez A, Mintardjo R, et al. Antibody and T-cell responses to a virosomal adjuvanted H9N2 avian influenza vaccine: impact of distinct additional adjuvants. *Vaccine* **2008**; 26:3640–6.
11. Kanekiyo M, Wei CJ, Yassine HM, et al. Self-assembling influenza nanoparticle vaccines elicit broadly neutralizing H1N1 antibodies. *Nature* **2013**; 499:102–6.
12. Limberis MP, Adam VS, Wong G, et al. Intranasal antibody gene transfer in mice and ferrets elicits broad protection against pandemic influenza. *Sci Transl Med* **2013**; 5:187ra72.
13. Lillie PJ, Berthoud TK, Powell TJ, et al. Preliminary assessment of the efficacy of a T-cell-based influenza vaccine, MVA-NP+M1, in humans. *Clin Infect Dis* **2012**; 55:19–25.
14. Weinfurter JT, Brunner K, Capuano SV III, et al. Cross-reactive T cells are involved in rapid clearance of 2009 pandemic H1N1 influenza virus in nonhuman primates. *PLoS Pathog* **2011**; 7:e1002381.